

BREWING
WITHOUT THE BLINDFOLD™

BSI



Yeast Supplier & Microbiology Lab

**BREWER'S
LABORATORY
HANDBOOK**

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“Brewing without the blindfold” means keeping an eye on your microbes, be they brewing yeast or the myriad contaminants that plague breweries. This handbook helps you do just that, and respects the fact that you want to spend time in the brewery, not in the lab!

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ASEPTIC TECHNIQUE

The ability to transfer cultures without contamination is imperative to good microbiology. Keep in mind:

- ❑ Hands must be washed and work areas must be cleaned with a 5% solution of disinfectant.
- ❑ Sterile vessels and liquids must be used (subject to 15# pressure for 15 minutes in a pressure cooker).
- ❑ Sterile transfer tools must be used (dip in 70% ethanol or rubbing alcohol and flame).
- ❑ Aerial fall-out and drafts must be minimized.

For More Information See Protocols:

- ❑ Pouring Plates
- ❑ Drawing Samples
- ❑ Swabbing
- ❑ Plating Samples Directly
- ❑ Plating Samples Using Filtration

QUICK and DIRTY “FILM TEST” FOR BOTTLED BEER

Bottled products can be checked for gross contamination without much difficulty.

The following are undesirable:

- ❑ Bottle necks with rings at the beer/headspace interface.
- ❑ Cloudiness in any part of the body.
- ❑ Residue at the bottom of filtered beer.
- ❑ Excessive foaming of opened, chilled beer.

Odor and taste are indicators as well. However, what is most useful is catching contamination well before it reaches taste, odor and visibility thresholds.

MICROSCOPIC EXAMINATION

For YEAST, use the 40x lens and make note of:

- ❑ Percent viability

For BACTERIA, use the 100x oil immersion lens and make note of:

- ❑ Whether cells are rods (bacilli) or rounds (cocci).
- ❑ The approximate length or diameter in microns.
- ❑ The gram reaction.

Microscopic examination of samples has limited use. The microscope is not very helpful for detecting even moderate levels of contamination, whether it be in yeast, wort or beer. If enough bacteria or wild yeast is present to be seen while surveying only a few fields, contamination levels are very high. Also keep in mind that different yeast strains can have very similar appearances under the microscope, so it can be impossible to tell a wild strain from a brewing strain, or to tell one brewing strain from another. Petite mutants are so-named because of their small colony size. They cannot be differentiated from normal cells under the microscope.

DETECTING MIXED STRAINS OF BREWING YEAST

If you use more than one strain of yeast in the brewery, you may wish to determine whether or not they are becoming intermingled. By plating the slurry for single colonies on wort agar and incubating for 5-7 days, you will be able to tell one from another after careful examination of colony shapes, sizes and colors. Any differences in these characteristics are solid indications of different strains.

For More Information See Protocols:

- ☐ Pouring Plates
- ☐ Streaking for Single Colonies
- ☐ Strain Purification

DETECTING BACTERIA and WILD YEAST with SELECTIVE MEDIA

If you know what type of contaminant you're encountering, you are better equipped to determine how serious a problem it presents and how to get rid of it. On a non-selective medium, such as wort agar, yeast colonies are relatively large, opaque, whitish and waxy-looking, while bacteria colonies are generally smaller, more translucent and slick-looking. Knowing this is fine, but a selective medium is needed for more specific identification. We recommend the following selective media for the basic groups of brewing contaminants:

- ☐ Brewing Yeast Bacteria - LMDA/SDA
- ☐ Wild Yeast - LCSM and LWYM

Each of these media are easy-to-use and allow for enumeration (counting the number of contaminants in the sample tested) and differentiation (knowing one type of organism on the plate from another), which is more than can be said for most testing media.

For More Information See Protocols:

- ☐ Pouring Plates
- ☐ Plating Samples Directly
- ☐ Plating Samples Using Filtration
- ☐ Streaking for Single Colonies
- ☐ Strain Purification

SIMPLE TESTS FOR IDENTIFYING BACTERIA

Make the following observations of any bacteria colonies before disturbing the colony with stain or other tests:

- ❑ Odor (vinegary, rotten, sulfury, fruity)
- ❑ Acid Production (halo surrounding colony with color change and/or clearing of medium)
- ❑ Colony Color, Size, Texture, Shape

The gram stain takes advantage of certain differences in cell membrane properties of bacteria. All bacteria are divided into two categories: gram-positive (takes on crystal violet stain and turns blue) and gram-negative (takes on safranin stain and turns pink). Yeasts are gram-positive. Armed with all the above information, check Table of Brewing Bacteria for bacteria's identity.

For More Information See Protocols:

- ☐ Gram Staining and Protocol.
- ☐ Catalase & Oxidase Tests.

CELL COUNTS and DETERMINATION of PROPER PITCHING RATE

A hemacytometer is essential for accurate cell counts (determination of number of yeast cells/ml in a slurry). The number of cells counted within a chosen grid is converted into cells/ml. This number is compared to the number of cells needed for healthy fermentation.

For More Information See Protocols:

- ☐ Cell Counting

PROTOCOLS

SAMPLE	FREQUENCY	SAMPLE SIZE	COMMON CONTAMINANTS	TOLERANCE
Water Supply	Once per Week	100ml, Filtered	Enteric, Molds	<10 cfu
Wort	Every Brew	1.0ml	Enteric, Acetic & Lactic, Wild Yeast	<10 cfu; 0 cfu wild yeast
Pitching Yeast	Every Crop	1.0ml	Enteric, Acetic & Lactic, Wild Yeast	<10 cfu; 0 cfu wild yeast
Fermenting Beer, days 1-2	Every Tank	1.0ml	Enteric, Acetic & Lactic, Wild Yeast	<10 cfu; 0 cfu wild yeast
Fermenting Beer, days 3-5	Every Tank	1.0ml	Acetic & Lactic, Wild Yeast	<10 cfu; 0 cfu wild yeast
Storage Tank	3 Times per Week	1.0ml	Acetic & Lactic	<10 cfu
Finishing Tank	3 Times per Week	1.0ml	Lactic	<10 cfu
Bottling Tank	Once per Month	100ml, Filtered	Lactic	<10 cfu
Bottled Beer	Every Batch	100ml, Filtered	Acetic & Lactic	<10 cfu
CIP'd Surfaces	Every CIP	Swab	-	0 cfu

Note: CFU = colony-forming units, or the number of colonies growing on the test plate.

PROTOCOL: DRAWING SAMPLES

Equipment

- 70% ethanol or rubbing alcohol
- Sterile sample tube

Procedure

- Wash and dry hands thoroughly.
- Sterilize intervening "outside" surfaces, such as valves, with 70% ethanol and flame.
- Do not uncap tube until immediately before sampling.
- Do not touch inside of tube or cap to any surface.
- If possible, allow sample to flow or crumble directly into tube without using utensils.
- Do not allow sample to overflow onto outside of tube.
- Recap tube as soon as possible.

PROTOCOL: SWABBING

Equipment

- ☐ Sterile swab tube

Procedure

- ☐ Wash and dry hands thoroughly.
- ☐ Unscrew swab/cap from tube without touching inside of tube or cap to any surface.
- ☐ Firmly swab area to be tested, twisting swab to expose entire tip to area in question.
- ☐ Immediately place exposed swab back into tube, tighten cap and label.

Notes:

Exposed swabs can be tested by streaking across the surface of plated media, or by adding 10ml sterile wort to swab tube and allowing to grow in a warm area (~86°F/30°C) for 3 days.

For More Information See Protocols:

- ☐ Plating Samples Directly
- ☐ Wort Stability Test

PROTOCOL: WORT STABILITY TEST

Equipment

- ☐ 70% ethanol or rubbing alcohol
- ☐ Sterile sample tube

Procedure

- ☐ Wash and dry hands thoroughly.
- ☐ Sterilize intervening “outside” surfaces, such as valves, with 70% ethanol and flame.
- ☐ Do not uncap sample tube until immediately before sampling.
- ☐ Do not touch inside of tube or cap to any surface.
- ☐ If possible, allow cooled, aerated wort to flow directly into tube without using utensils; fill tube about halfway.
- ☐ Do not allow sample to overflow onto outside of tube; recap loosely.
- ☐ Allow to grow in a warm area (~86°F/30°C) for 3 days.

Notes:

This test determines whether or not brewing organisms are present without allowing for their identification. Clear, bubble-free wort after 3 days of incubation at room temperature indicates the sample contains no viable brewing organisms. Cloudiness, bubbles or gas escaping when lid is loosened indicates that live organisms are present, which means either your sanitizing regime is inadequate, or sterile materials and surfaces are being exposed and contaminated.

PROTOCOL: POURING PLATES

Equipment

- ❑ 5% disinfecting solution
- ❑ Sterile petri dishes
- ❑ Sterile prepared media
- ❑ Permanent marker

Procedure

- ❑ Prepare media according to instructions.
- ❑ Wash and dry hands thoroughly.
- ❑ Clean work area with a 5% disinfecting solution such as Lysol or bleach.
- ❑ Lay out, label and date underside of plates and turn each back over, covers up.
- ❑ Allow media to cool until it is comfortably-warm to the touch (about 50°C).
- ❑ Swirl media occasionally to keep any insoluble components of media suspended.
- ❑ Pour 15ml (7ml for small plates) directly into each plate (about 5mm thick), lifting covers just enough to allow for pouring.
- ❑ Do not disturb plates until media has solidified.
- ❑ Invert and store plates refrigerated in a clean plastic bag.

Notes:

Pouring media while too hot will result in condensate inside the plate, which increases the chances of contamination. You can check plates for contamination by examining them after 48 hours of incubation in a warm area (~86°F/30°C). Wort agar (instead of commercial nutrient agar) = 30ml house wort, 70ml water, 2g agar; sterilize for 15 minutes at 15psi.

PROTOCOL: PLATING SAMPLES DIRECTLY

Plated media are generally used to test the purity of liquid samples such as wort, yeast slurry and bottled beer. It is important that the plates do not become contaminated with anything other than the sample you want tested, so store your plates refrigerated and upside-down in a clean plastic box until you are ready to use them.

Equipment

- ❑ Starsan or a 5% Lysol or bleach solution
- ❑ Media plates
- ❑ Sterile pipettes
- ❑ Plastic box

Procedure

- ❑ Choose an area that is enclosed or away from drafts and messy or dusty operations.
- ❑ Wipe down countertop or table with Starsan or a 5% Lysol or bleach solution.
- ❑ Wash and dry hands thoroughly.
- ❑ Inspect each new plate for contamination; label underside with date, sample name, sample site and size of sample plated.
 - » *Liquid sample*: place sample on media using a sterile pipette, gently spread over entire surface without gouging media and immediately replace plate lid.
 - » *Swab sample*: streak swab tip over entire media, twisting swab to expose all of tip to media surface and immediately replace plate lid.
- ❑ Place plates in box upside-down; loosely fit box lid and allow to grow in a warm area (~86°F/30°C) for 3 days.
- ❑ Do not wrap plates or seal plates in any way.

PROTOCOL: PLATING SAMPLES USING FILTRATION

If the entire sample is too large to be plated directly (>1.0ml), sample filtration is necessary. The purpose of filtration is to trap any organisms present onto a membrane filter by drawing the sample through it. The filter is then laid directly onto plated media.

Equipment

- Starsan or a 5% Lysol or bleach solution
- 70% alcohol
- Gas burner
- Vacuum pump
- Sterile filtration apparatus
- Sterile 0.45 μ m filters
- Tweezers
- Media plates

Procedure

- Choose an area that is enclosed or away from drafts and messy or dusty operations.
- Wipe down countertop or table with Starsan or a 5% Lysol or bleach solution.
- Wash and dry hands thoroughly.
- Inspect each new plate for contamination; label underside with date, sample name and sample site.
- Dip tweezers in alcohol and light with flame. Allow flame to extinguish on its own.
- Use tweezers to place sterile filter into filtration apparatus; reassemble apparatus.
- Flame mouth of sample vessel and add 100ml to the top receptacle.
- Cap top receptacle and attach vacuum to bottom receptacle. Draw entire sample through filter.
- Sterilize tweezers as above; remove top receptacle and retrieve filter¹.
- Place filter face-up on agar, making sure membrane's entire underside is in contact with media.
- To filter another sample, liberally spray alcohol into apparatus and draw through with vacuum; load with sterile filter.
- Wipe down a plastic box with Starsan or a 5% Lysol or bleach solution.
- Place plates in box upside-down; loosely fit box lid and allow to grow in a warm area (~86°F/30°C) for 3 days.
- Do not wrap plates or seal plates in any way.

Notes:

¹If another sample is to be filtered, disassemble apparatus and liberally spray down interior of top receptacle with alcohol; empty using vacuum suction. Sterilize tweezers, put fresh filter in place and repeat filtration procedure.

PROTOCOL: CATALASE & OXIDASE TESTS

Equipment

- 3% hydrogen peroxide
- Glass slide
- Oxidase "dry slides"

Procedure: Catalase Test

Drop a little hydrogen peroxide onto the colony in question, or onto bacteria smeared on a clean glass slide. A positive result is indicated by the formation of bubbles.

Procedure: Oxidase Test

Smear a little of the colony in question onto the slide.

A positive result is indicated by the bacteria turning a very dark "royal" purple color within 2 minutes.

PROTOCOL: GRAM STAINING

Equipment

- ☐ Microscope with 100x objective
- ☐ Immersion oil
- ☐ Glass slide
- ☐ Gram stain kit

Procedure:

- ☐ Place a drop of water on a clean glass slide and barely touch a flamed and cooled inoculating loop to the colony¹
- ☐ “Wash” loop in drop of water and evenly smear the suspension around in a pea-sized area; allow to air-dry.
- ☐ Hold glass slide about 3 inches above flame and hold for 4 seconds to kill and fix organisms to slide; allow slide to cool.
- ☐ Cover smear with crystal violet stain and leave on for 1 minute.
- ☐ Rinse and flood slide with iodine mordant; leave on for 3 minutes then drain.
- ☐ Slowly drip decolorizing rinse onto smear while holding slide vertically; stop as soon as color ceasing flowing from smear²
- ☐ Cover smear with safranin counterstain, leave on for 1 minute, then rinse gently with water and allow to air dry.
- ☐ Examine under microscope using oil-immersion lens. Gram⁺ is blue or purple; gram⁻ is red or pink.

Notes:

¹Adding too much bacteria to the water droplet will lead to too dense a smear, making good staining almost impossible. The proper amount of bacteria will be a barely visible dot of material on the loop.

²The decolorizing rinse is the most critical step. If you continually get gram⁻ or variable results for known gram⁺ bacteria, you are over-rinsing. Gram⁺ results for gram⁻ bacteria can result from too dense a smear.

PROTOCOL: STREAK-PLATING FOR SINGLE COLONIES

Equipment

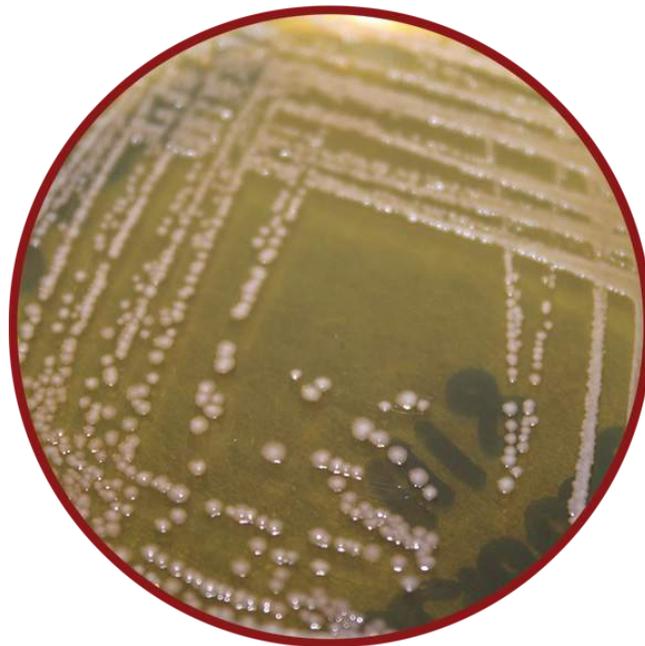
- ❑ Sterile water (contact-lens saline works well)
- ❑ Wort agar¹ plate
- ❑ Yeast culture

Procedure:

- ❑ Put three drops of sterile water next to each other on the periphery of a wort agar plate.
- ❑ Get a film of sample across an inoculating loop (for yeast slurry) or barely touch loop to a colony (for solid culture).
- ❑ Dip loop into first water drop and “wash” around until material from loop is suspended in water drop.
- ❑ Flame and cool inoculating loop.
- ❑ Get “film” from first drop across loop and transfer to second water drop, “washing” loop as before.
- ❑ Flame and cool inoculating loop.
- ❑ Get “film” from second drop across loop and transfer to third water drop, “washing” loop as before.
- ❑ Flame and cool inoculating loop.
- ❑ Get “film” from third drop across loop and streak across surface of plate, avoiding the three drops.

Notes

¹Wort agar = 30ml house wort, 70ml water, 2g agar; sterilize for 15 minutes at 15psi.



PROTOCOL: YEAST HARVEST & STORAGE

Equipment

- ❑ 70% ethanol or rubbing alcohol
- ❑ Sanitizer
- ❑ Storage container
- ❑ Cold water
- ❑ Sterile Wort

Procedure:

- ❑ Sterilize valves with 70% ethanol and flame, and sanitize any hoses, utensils and storage containers to be used.
- ❑ Catch the middle layer of yeast in a storage container as follows, discarding top and bottom layers:
- ❑ CONICAL vessels require draining the bottom trub-filled layer until good color and consistency is seen.
- ❑ ROUND vessels require manual harvesting with a paddle (layers mix if allowed to drain through the valve).
- ❑ Acidify about 100ml of water to ~pH 3 using a food-grade acid.
- ❑ For every gallon of yeast to be treated, add 2ml sodium chlorite¹ to the acidified water.
- ❑ When concoction turns a very pale yellow (at about 15 minutes) add to the yeast slurry, mixing well.
- ❑ Allow to sit for a minimum of 30 minutes (longer reaction time ensures effectiveness).
- ❑ Use yeast as needed or add an equal volume of sterile wort and store at 34°F/1°C with pressure relief for up to 2 weeks.
- ❑ If stored, test for viability, cell count and contamination before pitching.



Notes:

It is best to harvest yeast immediately after final gravity has been reached, before diacetyl rest. Even if it is kept very cold, yeast should not be stored in the cone or under beer. Finished beer is a nutrient-poor waste product to yeast, and forcing it to reside there will cause the viability to drop precipitously (over 50% within 48 hours), no matter how cold it is kept!

¹DioxyChlor™ from BIRKO at (800) 525-0476, or Star-Xene™ from Five Star at (800) 782-7019.

PROTOCOL: CELL COUNTING

Equipment

- Microscope with 40x objective
- Methylene blue, 1% aqueous
- Hemacytometer and cover slip, clean and dry

Staining

- Obtain a few mls of a slightly-cloudy suspension of yeast, keeping track of any dilutions¹ made.
- Dye with methylene blue dye, one drop at a time, until slurry is a cobalt blue.
- Shake sample until color is uniform; allow to sit for 1 minute.
- Place sample in one of the chambers using instructions that came with your hemacytometer.
- View under microscope using 40x objective; blue cells are non-viable, clear are viable.

Counting:

- Locate the central square with 25 smaller squares which are gridded in a 4x4 pattern.
- Count the total number of viable yeast in 5 of the squares², using pattern shown at page bottom.

Calculating Pitch Rate:

HOW MUCH YEAST DO YOU HAVE?

Slurry Density: _____ cells counted in 5 squares x 50,000 x _____ dilution factor¹ = _____ million cells/ml

Slurry Volume: _____ gal x 3800ml/gal = _____ ml

Total Cells Available: _____ million cells/ml x _____ ml = _____ Cells Available

HOW MUCH WORT DO YOU HAVE?

Pitch Rate: _____ °P x 1 million cells/ml = _____ million cells/ml

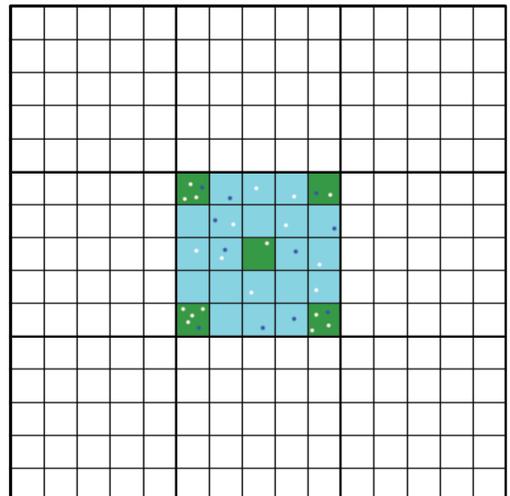
Wort Volume: _____ bbl x 31 gal/bbl x 3800ml/gal = _____ ml

Total Cells Required: _____ million cells/ml x _____ ml = _____ Cells Required

Notes:

¹Confused about making dilutions and calculating dilution factors? Consider a 1:10 dilution. The number on the left side of the colon is the volume (in ml, let's say) you begin with; the number on the right is the volume you end up with. A dense slurry may require a 1:100 dilution, namely, the addition of 1ml yeast to 99ml water, in order to render it countable. There is another way to get a 1:100 dilution. You may add 1ml yeast to 9ml water, mix, then remove 1ml of that suspension and add it to 9ml of fresh water. In other words, two 1:10 dilutions made in series are equivalent to a single 1:100 dilution. Similarly, three 1:10 dilutions made in series are equivalent to a single 1:1000 dilution. The dilution factor is the product of the numbers on right side of the colon. For example, a 1:10 dilution followed by a 1:4 dilution is equal to a 1:40 dilution and has a dilution factor of 40.

²Exclude buds and yeast lying on any two of the four borders of each of the 5 squares.



BACTERIA TYPES OVERVIEW

ACETIC ACID BACTERIA *Acetobacter, Acidomonas*

- From plant material
- Found in wort and sometimes bottled beer
- Contributes vinegary or cidery flavor

ENTERIC BACTERIA

Citrobacter, Enterobacter, Hafnia, Klebsiella, Obesumbacterium

- From water, soil, plant material
- Found in wort, early stages of fermentation
- Enterobacter and Citrobacter killed at pH 4.4 and 2% alcohol, others are not
- Contribute fusel alcohols, sulfur compounds, phenolics, and diacetyl

LACTIC ACID BACTERIA *Lactobacillus, Pediococcus*

- From grain dust
- Found throughout brewing process
- Contributes tart or sour flavor, diacetyl

NON-BREWING BACTERIA *Staphylococcus, Micrococcus, and Streptococcus*

- From skin cells, hair cells, dust, air, malt, saliva
- Not considered brewing bacterias
- May survive in beer, but does not produce off flavors

OTHER BACTERIA

Megasphaera, Pectinatus, Zymomonas

- Occur rarely
- Sources vary
- Found in areas with little oxygen, such as CO₂ recovery systems
- Zymomonas tolerates up to 10% alcohol
- Contributes sulfur compounds, acetaldehyde

BACTERIA IDENTIFICATION CHART

GENUS	GRAM	ACID	CATALASE	OXIDASE
Acetic Acid Bacteria (smell sharp and vinegary)				
Acetobacter 	-	+	+	-
Acidomonas 	-	+	+	+
Enteric Bacteria (smell rotten, sulfury, or like a bad cold)				
Citrobacter	-	-	+	-
Enterobacter	-	-	+	-
Haflnia	-	-	+	-
Klebsiella	-	-	+	-
Obesumbacterium	-	-	+	+
Lactic Acid Bacteria (smell sharp and fruity)				
Lactobacillus 	+	+	-	-
Pediococcus 	+	+	-	-
Other Bacteria				
Megasphaera 	-	+	+	-
Pectinatus 	-	-	-	-
Zymomonas 	-	-	+	-
Non-Brewing Bacteria				
Staphylococcus 	+	+	+	-
Streptococcus 	+	-	-	-
Micrococcus 	+	-	+	-

QUICK REFERENCE

Wort Aeration:

Aeration rates vary with wort volume, wort gravity and knockout time. In general, 8-10ppm of dissolved oxygen is desired for normal gravity worts. If aerating in-line with pure oxygen for the duration of a 1-hour knockout, this translates into 1.4LPM for every 10bbl of wort:

- ❑ **10ppm = 10mg/L = 11,780mg/10bbl**
- ❑ **1.4mg O₂ = 1ml O₂**
- ❑ **11,780mg O₂ = 8,414ml O₂ = 8.4L O₂**
- ❑ **8.4L of O₂ delivered over 1 hour = 0.14L delivered over 1 minute**

Sterilizing Time vs Temperature:

The system (not the liquid running through it!) must be kept at:

- ❑ **160°F (71°C) for 45 minutes**
- ❑ **170°F (77°C) for 30 minutes**
- ❑ **180°F (82°C) for 25 minutes**

Conversions

- ❑ **l = liter**
- ❑ **hl = hectoliter**
- ❑ **bbl = beer barrel**
- ❑ **3.8 l / gal**
- ❑ **100 l / hl**
- ❑ **118 l / bbl**
- ❑ **31 gal / bbl**
- ❑ **°C = 5/9 (°F - 32)**
- ❑ **°F = (9/5 °C) + 32**
- ❑ **° Plato = % sugar = ¼ (last two digits of specific gravity reading)**
- ❑ **5° Plato = 5% sugar = 1.020 sg**

OE = original extract, ° Plato **AE** - apparent extract , ° Plato **RE** = real extract, ° Plato

- ❑ **% alcohol by weight = 0.42 (OE - AE)**
- ❑ **% alcohol by weight = 0.52 (OE - RE)**
- ❑ **% alcohol by weight = 2.22 (RE - AE)**
- ❑ **% alcohol by volume = (% alcohol by weight x specific gravity of beer) / 0.791**

GLOSSARY

Aerobe: an organism which needs oxygen to survive (obligate aerobes will die without air).

Anaerobe: an organism which does not need air to survive (obligate anaerobes will die if exposed to air, while facultative anaerobes can grow with or without air).

Aseptic: against or without infection.

Contaminant: a relative term referring to any organism which lends undesirable characteristics; what is “bad” in one kind of brew may be intentionally cultured in another.

Inoculate: to introduce an organism to a sterile environment, intentionally or not.

Morphology: shape or appearance.

Petite mutant: a yeast with deficient respiratory abilities, resulting in slow growth and small (petite) colony size.

Strains: subspecies or “races” of *Saccharomyces cerevisiae*; all brewing yeasts are strains of this species.

Viability: percent of live cells in a population; not to be confused with vitality, which refers to metabolic activity or vigor of live cells.

Wild yeast: undesirable, undomesticated, or non-brewing yeast.

LAB SUPPLIES

ITEM	SIZE/QUANTITY	COMPANY
Wort Stability Test Jar	500ml	Cynmar
2 liter PP bottles	2000ml	Cynmar
Hemocytometer		Cynmar
Gram Stain Kit	each	Cynmar
Cotton Tip Applicators	box of 1000	Cynmar
Inoculating Loop	each	Cynmar
Sterile Transfer Pipettes	box of 500	Cynmar
Methylene Blue (1%)	each	Cynmar
Filter Apparatus	250ml	Fisher
Filter Apparatus	500ml	Fisher
Filter Apparatus	1000ml	Fisher
Filters/Filter App (0.45um)	box of 100	VWR
Hand Vacuum Pump	each	Cynmar
Sanitizer Spray Bottle	each	Cynmar
Forceps	each	Cynmar
Propane Torch	each	Hardware Store
Glass Slides	box	Cynmar
Immersion Oil	each	Cynmar
PP Samples Bottles	500ml	Cynmar
Anaerobic GasPak	box	Bacterius Ltd
New Monocular Microscope	each	Cynmar
New Binocular Microscope	each	Cynmar
Universal Beer Agar		Cynmar
70% Isopropyl Alcohol	each	Drugstore
3% Hydrogen Peroxide	each	Drugstore
Tupperware Box for Agar Plate Storage		
Tupperware Box for Incubator		

Fisher Scientific 1-800-766-7000

Cynmar 1-800-223-3517

VWR Labshop 1-888-897-5463

Bacterius Limited 1-832-474-4110*

*(additional charges for telephone orders may apply) sales@bacteria.us.com